

PROGETTO PLS

Matematica e Medicina: CELLULAR POTTS MODEL

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INTRODUCTION



All biological phenomena emerge from intricate interactions between multiple levels of organization:



INTRODUCTION



Mathematical approaches for biological problems employ a wide range of techniques, depending on the scale of interest:

| microscopic scale | mesoscopic scale | macroscopic scale | | |
|--|--------------------------------------|---|--|--|
| << 10 ⁻⁷ m subcellular level | 10 ⁻⁶ m cellular level | >> 10 ⁻⁵ m tissue level | | |
| RD SYSTEMS | INDIVIDUAL BASED MODELS | CONTINUOUS METHODS | | |
| ✓ systems of ODEs or PDEs | ✓ discrete ✓ phenomenologic | ✓ systems of PDEs ✓ populations as densities | | |
| ✓ kinetics equations | ✓ object-oriented | ✓ balance laws | | |

However, the use of a single specific type of model may be often unsatisfactory



The CELLULAR POTTS MODEL is:

- ✓ a hybrid and flexible individual-based approach, focused on the phenomenology of cell-level individuals (cells, ECM fibers, unicellular organisms,...)
- ✓ formed by a list of discrete individuals with phenomenological rules for their dynamics and interactions
- ✓ a Monte Carlo iterative method, based on an energy-minimization philosophy driving how the simulated individuals behave



The Cellular Potts Model (CPM) is a lattice-based Monte Carlo technique which follows an iterative and stochastic energy-minimization philosophy

a CPM domain is a d-dimensional lattice (i.e., a regular grid formed by identical d-dimensional lattice sites <u>x</u>), where d=2,3. Each site is identified by an integer number, called spin, $\sigma(x)$

a set of contiguous lattice sites labeled by the same spin σ form a single object, a discrete physical unit Σ_{σ} with a relative type $\tau(\Sigma_{\sigma})$

connections between neighboring lattice sites of unlike state σ represent objects' membranes

| 4 | 4 | 4 | 4 | 0 | 0 | 0 |
|---|---|---|---|---|---|---|
| 4 | 2 | 2 | 2 | 0 | 0 | 0 |
| 1 | 2 | 2 | 3 | 3 | 3 | 0 |
| 1 | 1 | 3 | 3 | 3 | 3 | 0 |
| 1 | 1 | 3 | 3 | 3 | 5 | 5 |
| 6 | 6 | 3 | 3 | 5 | 5 | 5 |
| 6 | 6 | 3 | 3 | 5 | 5 | 5 |



Grid subdomains may represent entire single biological elements





cells





matrix fibers

....or element subcompartments, for example their intracellular compartments (nucleus, cytosol, PM, Golgi Apparatus, ER, ...) and organelles (mitochondria, ...)





However, the more detailed is the cell representation, the more the model is computationally expensive.



The system energy is defined with an Hamiltonian H, which consists in the sum of terms relative to:

✓ adhesion between individuals (Steinberg's DAH)

 $H_{adh} = \sum_{x,x'} J_{\sigma(x),\sigma(x')} \left(1 - \delta_{\sigma(x),\sigma(x')}\right)$

✓ individual attributes (volume, surface, velocity,...)

 $H_{attr} = \sum_{i\text{-attribute},\sigma} [\lambda_{\sigma}^{i} (a_{\sigma}^{i} (t. v.) - A_{\sigma}^{i} (a. v.))^{2}]$

✓ effective and generalized forces (potential, chemotaxis, ...)

 $H_{force} = - \sum_{k-force,\sigma} \left[\mu^k_{\sigma} F^k_{\sigma} \right]$

 $J_{\sigma(x),\sigma\,(x')},\lambda_{\sigma}$ and μ_{σ} are Potts coefficients describing the importance of the relative biophysical properties or mechanisms



Individuals move and behave in order to iteratively and stochastically reduce H, with a simple algorithm:

1- choose a lattice site belonging to an individual membrane and attempt to copy its spin into a randomly chosen neighboring lattice site



2- the difference in the system energy as a results of the attempt is calculated: $\Delta H = H_{after \ spin \ copy} - H_{before \ spin \ copy}$

3- the attempt is accepted with a Boltzmann-like probability:

 $P(\Delta H) = p(T_{\text{moving object }\Sigma\sigma})\min\{1, \exp(-\Delta H / T_{\text{moving object }\Sigma\sigma})\},\$

where

 $T_{moving object \Sigma\sigma}$ is the agitation rate of the object moving site belongs to

- p is a maximum transition probability function characterized by:
 - p(0) = 0
 - $\lim_{T \to +\infty} p(T_{\text{moving object}}(t)) = 1$



4- the algorithm is repeated until the system reaches a global minimum or until a given observation time. Each iteration is defined a Monte Carlo Step



The model can be finally integrated by the evolution of molecular variables (i.e., ions, molecules, or genes) localized both within biological elements (within one of their subcompartments) and/or in the extracellular space. The dynamics of such microscopic variables are modelled by typical and suitable reaction-diffusion equations:

$$\frac{\partial c(x)}{\partial t} = D_c \partial^2 c / \partial x^2 - a c + s$$
variation of c
within point x at
time t

Finally, we need to define constitutive laws describing how the molecular elements influence cell behavior



For example:

 ✓ the intracellular level of active cadherins or integrins will influence the cell-cell ahesion energy

 ✓ The level of extracellular growth factor will influence cell motilty





OVARIAN CANCER TRANS-MESOTHELIAL INVASION (with Prof. A. Funaro)



MS, Giverso, Lo Buono, Preziosi, Funaro, Math Model Nat Phenom, 2010.



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WOUND HEALING ASSAY OF EPITHELIAL CELLS IN RESPONSE TO A MOTILITY FACTOR (i.e., HGF)





WOUND HEALING ASSAY OF EPITHELIAL CELLS IN RESPONSE TO A MOTILITY FACTOR (i.e., HGF)



t = 0 h



WOUND HEALING ASSAY OF EPITHELIAL CELLS IN RESPONSE TO A MOTILITY FACTOR (i.e., HGF)



t = 12 h



WOUND HEALING ASSAY OF EPITHELIAL CELLS IN RESPONSE TO A MOTILITY FACTOR (i.e., HGF)

the cell mass can be sorted into three subpopulations, namely internal, middle and external, characterized by well-defined migratory behavior









DIFFERENT MORPHOLOGIES OF TUMOR INVASION FRONTS



DIFFERENT MORPHOLOGIES OF TUMOR INVASION FRONTS



in silico and *in vivo* pT1 urothelial carcinoma invading into the lamina propria, with single aggressive malignant cells detaching from the main tumor mass

DIFFERENT MORPHOLOGIES OF TUMOR INVASION FRONTS



in silico and *in vivo* microinvasive tumor of the cervix, with fingers of invading cells protruding through the basement membrane



DIFFERENT MORPHOLOGIES OF TUMOR INVASION FRONTS



simulation of the evolution of a glioma spheroid. The tumor invasiveness is strictly regulated by intercellular adhesion



APPLICATIONS: CELL MIGRATION IN 3D SCAFFOLDS

in the presented model, cells are defined as compartmentalized individuals differentiated in two basic objects: the nucleus ($\tau = N$) and in the cytosolic region ($\tau = C$)

the extracellular environment is instead formed by a two-component matrix, where:

✓ matrix fibers (τ =F, yellow) are bidimensional basic objects with defined measures and topology;

✓ the interstitial medium (τ =M, black) is a generalized object isotropically distributed throughout the simulation domain



TO DE LORIZONI

APPLICATIONS

CELL MIGRATION IN 3D MATRIX SCAFFOLDS



cells are seeded in an isotropic two-component matrix scaffold, characterized by a regular mesh of inelastic fibers with square pores cell movement is Brownian with a mean net displacement of 75 μ m in 12 h, a velocity of 14 μ m/h , and a persistence time less than 1.5 h



CELL MIGRATION IN 3D MATRIX SCAFFOLDS

decreasing cell-fiber adhesion



a bimodal relation is found between cell motile behavor and cell-fiber adhesion

CELL MIGRATION IN 3D MATRIX SCAFFOLDS



the directional component of cell motion increases until, in the case of all fibers aligned along the x-axis, cells movement is almost linear, with no change in cell velocity.



CELL MIGRATION IN 3D MICROCHANNELS



bottom channel size < nucleus diameter < middle channel size < cell diameter < top channel size The model reproduces a microfabricated device with channels of various width and a planar surface just outside their entrance. The cell migratory behavior is characterized by one of the following categories: i) cells that only penetrate the channel with a part of their cytoplasm are classified as penetrating, ii) cells that completely enter in the channel structure but are not able to migrate to the other side within the observation period are called invasive, iii) cells that reach the opposite border of the channel are finally termed permeative.

CELL MIGRATION IN 3D MICROCHANNELS



Migratory behavior of cells with an elastic cytosol and a still rigid nucleus. They are now able to enter also in the intermediate channel, displaying a permeative behavior. However they are constrained to stay outside the smaller structure.

CELL MIGRATION IN 3D MICROCHANNELS



Migratory behavior of cells with an elastic cytosol and a deformable nucleus. The enhancement in nucleus elasticity enables cells to enter also in the smallest channel, where they acquire an invasive phenotype





Blood vessel formation is a complex and multilevel process fundamental in:



menstrual cycle

mammary gland during lactation

granulation tissue after wound healing

pathological situations

chronic inflammatory disease

vasculopathies

tissue injuring in ischemia

cancer progression

Vascularization is a pivotal step in tumor development, providing the necessary nutrients and allowing malignant cells enter in the circulatory system





The discovery of efficient antiangiogenic therapies is a fundamental issue in cancer research and treatment has given rise to multiple experiments, which aim to understand the key mechanisms involved in malignant vascularization and to identify intervention strategies potentially able to disrupt them



Experimental image of a vascularized solid tumor, courtesy of the Institute for Cancer Research and Treatment, Candiolo, Italy

(with Prof. L. Munaron)



The system of equations regulating the intracellular biochemical pathways is given as

Extracellular VEGF

$$\frac{\partial h_{\text{VEGF}}}{\partial t} = \begin{bmatrix} D_{\text{VEGF}} \partial^2 h_{\text{VEGF}} / \partial x^2 \\ \text{diffusion} \end{bmatrix} - \begin{bmatrix} k_{\text{VEGF}} h_{\text{VEGF}} \\ \text{decay} \end{bmatrix} - \begin{bmatrix} y_{\text{VEGF}} h_{\text{VEGF}} \\ \text{cell uptake} \end{bmatrix} + \begin{bmatrix} s_{\text{VEGF}} \\ \text{addition} \end{bmatrix}$$

Intracellular messengers
$$\frac{\partial h_{AA}}{\partial t} = \begin{bmatrix} D_{AA} \partial^2 h_{AA} / \partial x^2 \\ D_{AA} \partial^2 h_{AA} / \partial x^2 \end{bmatrix} - \begin{bmatrix} k_{AA} h_{AA} \\ + \end{bmatrix} + \begin{bmatrix} j_{AA} \text{VEGFR} \\ j_{NO} \text{VEGFR} \end{bmatrix} + \begin{bmatrix} c_{AA} \text{Ca} \\ c_{NO} \text{Ca} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ b_{AA} h_{AA} \end{bmatrix} + \begin{bmatrix} j_{NO} \text{VEGFR} \\ c_{NO} \text{Ca} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ b_{AA} h_{AA} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ c_{AA} \text{Ca} \\ c_{AA} \text{Ca} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ c_{AA} \text{Ca} \\ c_{AA} \text{Ca} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ c_{AA} \text{Ca} \\ c_{AA} \text{Ca} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ c_{AA} \text{Ca} \\ c_{AA} \text{Ca} \\ c_{AA} \text{Ca} \end{bmatrix} + \begin{bmatrix} b_{AA} h_{AA} \\ c_{AA} \text{Ca} \\ c_{A$$

Intracellular calcium







(A)





 $50 \text{ px} \approx 50 \text{ } \mu\text{m}$

(B)

(C)



(D)

(E)





Evolution of intracellular calcium level during tubulogenesis



calcium signals, which are typically peripheral restricted, are detectable in the initial phase of the process, while they are down regulated during the maturation of EC tubules. The initial increment of calcium levels is in fact necessary for the enhancement of cell migratory properties

Inhibition of calcium entry: carboxyamidotriazole,CAI, compound



Experimental image courtesy of LM and of the Department of Animal and Human Biology, Universita degli Studi di Torino.

complete disruption of tubule formation, as the TECs remain almost scattered

Exclusion of either AA or NO biosynthesis - AACOCF3 or L-NAME drugs



formation of immature networks, where several branches have partially formed, but have not been able to organize into a single structure



Blocking cysotkeletal remodeling: phalloidin-like compounds



formation of clumped, stunted, shorter and thicker sprouts, Disruption of cell persistent movement



formation of immature and swollen sprouts characterized by large intervascular spaces

Increasing VEGF degradation:

Disruption of chemotaxis



formation of a reduced-in-scale network



formation of poorly structured islands similar to those obtained by extinguish VEGF gradients



The model allows to prove the efficacy some anti-angiogenic therapies that are currently in use or in trial and to test the potential of other biomedical intervention strategies

| | EXISTING COMPOUND | P-VALUE (pct) | EFFICIENCY | | | |
|----------------------------|---------------------------------------|---------------|------------|----|--|--|
| VEGF UPTAKE | sorafenib, sunitinib, vatalanib | 0.00082 | +++ | | TECs remain scattered | |
| VEGF DEGRADATION | | 0.00234 | ++ | | reduced-in-scale pattern | |
| CALCIUM ENTRY | CAI | 0.00085 | +++ | | TECs remain scattered | |
| AA PRODUCTION | AACOCF3 | 0.00425 | + | | formation of immature networks | |
| NO PRODUCTION | L-NAME, L-NMMA | 0.00473 | + | JL | | |
| CYTOSKELETAL REMODELING | phalloidin | 0.00092 | +++ | | clumped sprouts with large intervascular spaces | |
| PERSISTENCE | | 0.00139 | ++ | | | |
| CHEMOTAXIS | | 0.00097 | +++ | | poorly structured islands | |
| ADHESION | anti-VE-cadherin antibodies | 0.00069 | +++ | | TECs remain scattered | |
| MARCO SCIANNA | | | | | | |

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